Notes

ethanol gave the corresponding mixture of Z alcohol 2 and its E isomer 9 in a quantitative yield. Aldehydes 3 and 4 could be separated⁹ prior to this step or alcohols 2 and 9 would have to be isolated after reduction.

Experimental Section

All boiling points are uncorrected. Infrared spectra were determined with a Perkin-Elmer Model 137 spectrometer. NMR spectra were determined in deuteriochloroform solution on Varian Associates spectrometers, Models A-60 or XL-100. Line positions are given in δ scale, with tetramethylsilane as an internal standard. Mass spectra were recorded on an Atlas CH-4B or Associated Electrical Industries MS-902 spectrometer, high resolution measurements being determined with the latter instrument.

3.3-Dimethylcyclohexanone Diethyl Ketal (7). 3.3-Dimethylcyclohexanone (6) (6.54 g, 51.0 mmol), triethyl orthoformate (8.98 g, 60.6 mmol), and p-toluenesulfonic acid (6 mg) in 38 mL of anhydrous ethanol were stirred at room temperature for 64 h. Enough ethanolic NaOEt was added to neutralize the reaction mixture and the latter was concentrated under vacuum. Fractional distillation through a spinning-band column gave 9 g (87%) of pure 3,3-dimethylcyclohexanone diethyl ketal (7): bp 82-83 °C (13 mm); IR (neat) 1385, 1359 (-C(CH₃)₂), 1195, 1174, 1119, 1093, 1058 cm⁻¹ (COCOC); ¹H NMR $(\text{CDCl}_3) \delta 0.95$ (s, 6 H, geminal CH₃), 1.14 (t, 6 H, J = 7 Hz, CH₃CH₂-), 1.49 (s, 2 H, (RO)₂CCH₂C(CH₃)₂), 3.45 (q, 4 H, J = 7 Hz, CH₃CH₂-); mol wt 200.1777 (calcd for C₁₂H₂₄O₂: 200.1776)

2-(1-Ethoxy-3,3-dimethylcyclohexyl)-1,1-diethoxyethane (8). In a three-necked flask equipped with a rubber septum and a dry ice condenser protected with a calcium chloride tube are placed 3,3dimethylcyclohexanone diethyl ketal (7) (7.37 g, 36.9 mmol) and 3 mL of a solution of 10% ZnCl₂ in ethyl acetate. The mixture was stirred and maintained at a temperature of 45 °C while 3.8 mL of ethyl vinyl ether (40.5 mmol) was added dropwise with a syringe. After the addition was completed the mixture was stirred for 2 h at 45-50 °C (bath temperature) and then allowed to come to room temperature. The mixture was diluted with ether and washed with a solution of 5% NaOH and the ethereal layer was dried over Na₂CO₃. Filtration and removal of the solvent gave 9.45 g (94%) of 2-(1-ethoxy-3,3-dimethylcyclohexyl)-1,1-diethoxyethane (8), which was purified by distillation using a spinning band column: bp 79-80 °C (0.5 mm); IR (neat) 2950, 1389, 1370, 1183, 1124, and 1064 cm $^{-1}$; ¹H NMR (CDCl₃) δ 0.86 and 1.05 (two singlets for geminal methyl group), 1.12 and 1.18 (t, 3 H, CH₃CH₂O), 1.74 (d, 2 H, J = 4 Hz, EtOCCH₂CH(OEt)₂), 3.2–38 $(m, 6 H, OCH_2CH_3), 4.68 (t, 1 H, J = 4 Hz, CH(OEt)_2).$

Preparation of (Z)-3,3-Dimethyl- $\Delta^{1,\alpha}$ -cyclohexaneacetaldehyde(3) and (E)-3,3-Dimethyl- $\Delta^{1,\alpha}$ -cyclohexaneacetaldehyde (4). To acetal 8 (950 mg, 3.4 mmol) dissolved in 10 mL of glacial acetic acid and 0.77 mL of water was added 1.1 g of sodium acetate. The reaction mixture was stirred and heated for 3 h at 95 °C under an atmosphere of nitrogen. After cooling the resulting mixture was poured into ice, basified by careful addition of solid NaHCO₃, and extracted with ether. The combined extracts were washed with water and 5% aqueous NaHCO₃ and dried over anhydrous Na₂CO₃. Filtration and removal of the solvent gave 450 mg (84%) of a mixture⁷ of aldehydes 3 and 4, showing properties consistent with those reported by Tumlinson:² IR (neat) 1681 and 1639 cm⁻¹; ¹H NMR (CDCl₃) δ 0.96 (s, 6 H, geminal CH₃). 2.24 (t, 2 H, $-CH_2-$ trans to -CHO), 2.50 (s, 2 H, $-CH_{2}$ - cis to -CHO), 5.92 (d, 1 H, J = 8 Hz, -C=CH-), and 9.98 (d, 1 H, J = 8 Hz, -CHO) were assigned to 3, while those peaks at 0.93 (s, 6 H geminal CH₃), 2.08 (s, 2 H, -CH₂- trans to -CHO), 2.68 (t, 2 H, -CH₂- cis to -CHO), 5.78 (d, 2 H, J = 8 Hz, -C=-CH-), and 10.02 (d, 1 H, J = 8 Hz, -C HO) were assigned to 4. NMR peaks at 1.2–1.9 were common to both isomers; mol wt 152.1206 (calcd for C10H16O: 152.1201).

Preparation of the Isomeric Alcohols (Z)-3,3-Dimethyl- $\Delta^{1,\beta}$ -cyclohexaneethanol (2) and (E)-3,3-Dimethyl-($\Delta^{1,\beta}$ -cyclohexaneethanol (9). A mixture of 400 mg of aldehydes 3 and 4 (6:4 ratio) in 20 mL of absolute ethanol and 400 mg of NaBH₄ was stirred at ambient temperature for 1 h. The reaction mixture was hydrolyzed with water and extracted with CH2Cl2. The combined extracts were washed with water and dried over anhydrous MgSO₄. Filtration and removal of the solvent gave the isomeric alcohols 2 and 9 in a quantitative yield, showing properties consistent with those reported by Tumlinson:² IR (neat) 3400, 1667, 1075, 1031, 1000 cm⁻¹; ¹H NMR (CDCl₃) δ 0.90 (s, 6 H, geminal CH₃), 1.92 (s, 2 H, -CH₂- cis to -CH₂OH), 2.07 (t, 2 H, -CH₂- trans to -CH₂OH), 4.12 (d, 2 H, J = 7 Hz, -CH₂OH), and 5.48 (t, 1 H, J = 7 Hz, -C=:CH₋) were assigned that there at 0.87 (a, 5 H, geminal CH₂) 1.80 (c, 2 H, -CH₂- trans to 2, while those at 0.87 (s, 6 H, geminal CH_3), 1.89 (s, 2 H, $-CH_2$ - trans to $-CH_2OH$), 2.12 (t, 2 H, $-CH_2$ - cis to $-CH_2OH$), 4.14 (d, 2 H, J = 7

Hz, $-CH_2OH$), and 5.32 (t, 1 H, J = 7 Hz, $-C=CH_-$) were assigned to 9. NMR peaks at 1.2-1.8 were common to both isomers; mol wt 154.1341 (calcd for $C_{10}H_{18}O$: 154.1357).

Acknowledgment. We would like to thank Professor James P. Kutney of the University of British Columbia, Vancouver, Canada and Dr. Paul Baker of CPPN, Rio de Janeiro, Brazil for spectral determinations. The partial support of this work by the National Research Council of Brazil and CAPES is gratefully acknowledged.

Registry No.-2, 26532-23-0; 3, 26532-24-1; 4, 26532-25-2; 6, 2979-19-3; 7, 65392-27-0; 8, 65392-28-1; 9, 30346-27-1; triethyl or-thoformate, 122-51-0; ethyl vinyl ether, 109-92-2.

References and Notes

- (1) M. Jacobson, "Insect Sex Attractants", Interscience, New York, N.Y., 1965.
- (a) J. H. Tumlinson, D. D. Hardee, R. C. Gueldner, A. C. Thompson, P. A. Hedin, and J. P. Minyard, *Science*, **166**, 1010 (1969); (b) J. H. Tumlinson, R. C. Gueldner, D. D. Hardee, A. C. Thompson, P. A. Hedin, and J. P. Minyard, (2)J. Org. Chem., 36, 2616 (1971).
 J. H. Babler and T. R. Mortell, *Tetrahedron Lett.*, 669 (1972)

- (3) J. H. Babler and T. R. Mortell, *Tetrahedron Lett.*, 669 (1972).
 (4) S. W. Pelletier and N. V. Mody, *J. Org. Chem.*, 41, 1069 (1976).
 (5) For a synthesis from geranic acid, see R. H. Bedoukian and J. Wolinsky, *J. Org. Chem.*, 40, 2154 (1975).
 (6) (a) G. Buchi, O. Jeger, and L. Ruzicka, *Helv. Chim. Acta*, 31, 241 (1948); (b) H. O. House and W. F. Fischer, *J. Org. Chem.*, 33, 949 (1968).
 (7) O. Isler, H. Lindlar, M. Montovan, R. Rüegg, and P. Zeller, *Helv. Chim. Acta*, 20 (2065).
- 39, 249 (1956).
- (8) The ratio 6:4 was determined via integration of the vinyl hydrogen NMR absorption band for each isomer.
- For a description to separate aldehydes 3 and 4, refer to J. H. Tumlinson, R. C. Guelder, D. D. Hardee, A. C. Thompson, P. A. Hedin, and J. P. Minyard, "Chemicals Controlling Insect Behavior", M. Beroza Ed., Academic Press, (9) New York, N.Y., 1970, p 50.

Cobalt-Catalyzed Oxidation of Isotopically Labeled Cyclohexanone

J. D. Druliner

Contribution No. 2512, Central Research and Development Department, E. I. du Pont de Nemours and Company, Wilmington, Delaware 19898

Received August 15, 1977

A large body of literature exists covering various aspects of metal-catalyzed oxidation of cyclohexanone or cyclohexane to yield predominantly adipic acid.¹⁻⁴ We report herein results of studies on cobalt-catalyzed oxidations of isotopically labeled cyclohexanone. Of the numerous by-products formed, glutaric and succinic acids are formed in 10-20% yields. We examined the fate of cyclohexanone labeled at the carbonyl carbon with ¹⁴C and with ¹³C to try to resolve conflicting mechanistic proposals for the formation of glutaric and succinic acids.

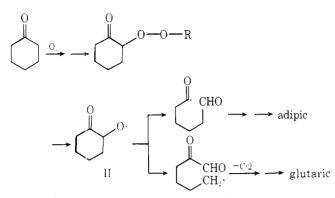
The percentage retention of the carbonyl carbon from labeled cyclohexanone in glutaric and succinic acid products was 91% in glutaric acid and 87% in succinic acid as shown in Table I.

The high retention of C-1 from labeled cyclohexanone is not consistent with a proposal that glutaric acid arises by exclusive

Table I. Oxidation of [1-14C]- and [1-13C]Cyclohexanone (K): Percent Retention of C-1 by Glutaric and Succinic Acids

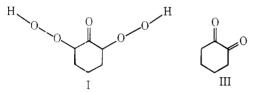
	% retention of C-1 Glutaric/ Succinic/		% K	% yields from K		
K	adipic	adipic	sion	Adipic	Glutaric	Succinic
[1- ¹⁴ C]	88	82	63	18	5	0.4
[1- ¹³ C]	96	84	62	23	4	0.5
[1- ¹³ C]	88	94	34	37	8	1.1
Average	91	87				

0022-3263/78/1943-2069\$01.00/0 © 1978 American Chemical Society



loss of C-1 from an intermediate 2,6-dihydroperoxycyclohexanone compound (I).⁵ A more recent proposal that glutaric acid is formed by loss of C-2 from an α -ketooxy radical (II) is consistent with the observed high retention of the labeled carbonyl carbon.8

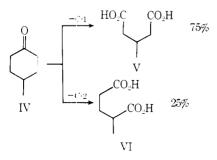
It has been reported that α -ketooxy radicals undergo thermolysis or metal-catalyzed cleavage to generate carboxylic acids in which the carboxyl group is derived from the keto group.⁶ It has been proposed that 1,2-cyclohexanedione (III)



is formed during oxidation of cyclohexanone and gives rise to most of the CO and CO₂ evolved.⁷ Although 1,2-cyclohexanedione may account for CO and CO_2 generation, it is not a viable intermediate for glutaric or succinic acid formation. Loss of a carbonyl carbon from III would decrease the isotopic enrichment by a factor of 2.

Significant glutaric acid formation from decarboxylation of adipic acid is not consistent with the observed high degree of retention of C-1 from labeled cyclohexanone or with reported carboxylic acid decarboxylation studies.^{8,9} The extent to which succinic acid is derived from solvent acetic acid was examined using [1-¹⁴C]acetic acid under the oxidation conditions used for labeled cyclohexanone. The conversion of cyclohexanone was 44% with a yield to adipic acid of 27%. Radiometric analysis of isolated succinic acid showed the presence of 0.04 mol of [1-14C]carboxyl groups per mole of succinic acid. Assuming all HO₂C--CH₂ radicals were derived from acetic acid, about 2% of the succinic acid was derived from acetic acid.

The effect of methyl substitution on the extent of loss of C-1 was found to be substantial. Whereas unsubstituted (labeled) cyclohexanone gave glutaric acid with >90% retention of C-1, 4-methylcyclohexanone (IV) gave a 3-methylglutaric (V)/ 2-methylglutaric (VI) acid ratio of 3:1. The stability of diacid products V and VI to reaction conditions was checked by replacing half of the cyclohexanone in a typical oxidation experiment with equal molar amounts of V and VI. Of the cyclohexanone charged, 34% was converted to adipic acid. The ratio of V/VI in the product was 1.0 within experimental error



based on ¹H NMR integrals for the methyl resonances for V $(\delta 1.05)$ and VI $(\delta 1.15)$. The preponderance of loss of C-1 with methyl substitution may be largely a steric effect.

Experimental Section

All oxidation reactions were carried out in 10-cm³ stainless steel shaker tubes at 100 °C with 200 psi of O₂ for 10-16 h. A typical starting solution consisted of 5.5 g (91.7 mmol) of acetic acid, 1.05 g (10.7 mmol) of [1-14C]cyclohexanone, and 0.10 g (0.42 mmol) of Co(OAc)₃. Gas chromatographic analyses for adipic, glutaric, and succinic acids were carried out on a 12 ft \times 0.12 in. stainless steel column of OV-1 at 190 °C following treatment with BSTFA to form trimethylsilyl diesters. [1-14C]Cyclohexanone was commercially available.¹⁰ [1-¹³C]Cyclohexanone was prepared by carbonylation of bisborinane with ¹³CO.¹¹ Products containing ¹⁴C were analyzed with a Packard liquid scintillation spectrometer using standard dilution techniques. Each sample was crystallized repeatedly to obtain constant specific activity. Products containing ¹³C were analyzed by gas chromatography/mass spectroscopic techniques.¹²

Acknowledgment. We thank Drs. B. A. Carlson, U. Klabunde, E. J. Lukosius, and J. B. Sieja for helpful discussions

Registry No.—Co(OAc)₃, 917-69-1; cyclohexanone, 108-94-1; glutaric acid, 110-94-1; succinic acid, 110-15-6.

References and Notes

- (1) I. V. Berezin, E. T. Denisov, and N. M. Emanuel, "The Oxidation of Cyclohexane", Pergamon Press, London, 1966. A. Onopchenko and J. G. D. Schulz, *J. Org. Chem.*, **38**, 3729 (1973); **40**,
- (2)3338 (1975)
- 3338 (1975).
 (3) D. G. Hendry, C. W. Gould, D. Schuetzle, M. G. Syz, and F. R. Mayo, *J. Org. Chem.*, **41**, 1 (1976).
 (4) K. Tanaka, *Prepr. Div. Pet. Chem. Am. Chem. Soc.*, **19**, 103 (1974).
 (5) G. N. Koshel et al., *Neftekhimiya*, **14** (2), 263 (1974).
 (6) Y. Sawaki and Y. Ogata, *J. Org. Chem.*, **41**, 2340 (1976).
 (7) I. I. Korsak and V. E. Agabekov, *Dokl. Akad. Nauk B. SSR*, **19** (8), 711 (1075).

- (1975) (a) (A), B. Tinker, J. Catal., 19, 237 (1970); (b) S. S. Lande and J. K. Kochi, J. Am. Chem. Soc., 90, 5196 (1968).
- Reaction product vapors contained too little CO and CO₂ for quantitative isotopic measurements. At 100 °C decarboxylation of solvent acetic acid (9) tocold account for a significant fraction of the total CO and CO₂. It is reported that, at 150 °C, ''about 30% of the total carbon oxides results from its (acetic acid) decomposition.'' ^{8a}
- (10)1-14C]Cyclohexanone was obtained from Amersham/Searle Corp. That [1-⁺C]Cyclonexanone was obtained from Ameristian Seare corp. That the ¹⁴C was contained only in C-1 was shown by oxidizing [1-¹⁴C]cyclo-hexanone to adipic acid and then converting the adipic acid to cyclopen-tanone by BaO-catalyzed decarboxylation. The specific activity of the cyclopentanone was exactly half that of the starting [1-¹⁴C[cyclohexa-
- (11) H. C. Brown, "Organic Syntheses Via Boranes", Wiley, New York, N.Y.,
- 1975, p 158. (12) GC/MS was done using chemical ionization by T. A. Blazer, Central Research and Development Department.

Synthesis of N-(4-Azido-2-nitrophenyl)amino-1-alkyl- β -D-glucopyranosides: Photoaffinity Labeling **Derivatives of Glucose**

Myrna Hagedorn, Ronald R. Sauers,* and Alexander Eichholz

Department of Chemistry, Rutgers-The State University of New Jersey, New Brunswick, New Jersey 08903, and The Department of Physiology, College of Medicine and Dentistry of New Jersey, Rutgers Medical School, Piscataway, New Jersey 08854

Received November 21, 1977

During the course of studies of the mechanism of glucose transport in human erythrocytes, the need arose for derivatives of glucose which could serve as photoaffinity labeling agents.^{1,2} Since an integral part of these studies involved the evaluation of reagents in which the distance between the sugar moiety and the photolabile grouping was varied systemati-

*Address correspondence to this author at Rutgers—The State University of New Jersev

0022-3263/78/1943-2070\$01.00/0 © 1978 American Chemical Society